

DIABETIC RETINOPATHY: NEW PHARMACOLOGICAL TARGETS

F. Lazzara

Department of Biomedical and Biotechnological Science, University of Catania, Catania, Italy

E-mail: francesca.lazzara@unict.it. ORCID: 0000-0002-9825-2104

Doi: 10.36118/pharmadvances.2022.27

SUMMARY

Diabetic retinopathy (DR) is a secondary complication of diabetes mellitus and represents the most common cause of irreversible vision loss in working people of industrialized countries. DR is generally considered a microvascular complication of diabetes, although the inflammatory component plays a key role. The main cause of vision loss in diabetic patients with proliferative diabetic retinopathy is the diabetic macular edema (DME), that is responsible to the retinal detachment. DME is mainly caused by new angiogenesis, which is a hallmark of the advanced stage of DR (proliferative diabetic retinopathy, PDR). Retinal neovascularization is principally driven by pro-angiogenic factors (e.g., VEGF-A, PlGF), inflammatory mediators (TNF- α , interleukins, chemokines) and oxidative stress-related elements. Chronic hyperglycemia is the primary causative factor of DR, however, several points of DR etiopathogenesis are still unclear. Many other factors are involved during the early stages of DR such as the retinal hypoxia, that is a trigger of VEGF release in the back of the eye. Up to now, the pharmacological approaches for DR are intravitreal anti-VEGF agents and corticosteroids. However, some patients can be refractory to anti-VEGF therapy, therefore, efforts must be carried out to discover novel pharmacological targets to handle DR. Hereby, the current literature will be revised about novel potential pharmacological targets, with a focus on PlGF, miRNAs and purinergic P2X7 receptor. Future drug development campaigns on these targets might lead to better clinical outcomes, possibly in the early phase of the disease.

Key words

Diabetic retinopathy; angiogenesis; hypoxia; anti-VEGF; inflammation.

Impact statement

Currently, management and treatment of diabetic retinopathy (DR) are characterized by several unmet medical needs. Particularly, early-stage DR lacks of approved therapeutical intervention, and its pathogenesis is multifactorial. Pharmacological research should focus on novel pharmacological targets, that address pathogenetic factors of DR, such as inflammation, oxidative stress and angiogenesis.

INTRODUCTION

Diabetic retinopathy (DR) represents a major public health concern, and it is the leading cause of vision loss in working age (1). The prevalence of DR among diabetic patients is about 40%, and approximately 5-10% of these individuals have vision threatening conditions (2, 3). Chronic hyperglycemia is the primary causative factor

of diabetic retinopathy, however, etiopathogenesis of DR is still unclear (4-7). Ophthalmologists classify diabetic retinopathy mainly into two stages, the non-proliferative diabetic retinopathy (NPDR) and proliferative diabetic retinopathy (PDR) (8). The NPDR is characterized by lesions due to chronic hyperglycemia, that can lead to microaneurysms due to instability of

capillary walls. As soon as microaneurysms start leaking, NPDR can develop in macular edema and consequent impaired vision, due to deposition of fluid under the macula. The presence of this fluid, composed by lipids, leads to the formation of yellow deposits, called hard exudates. Moreover, with the progression of the disease, the affected vessels can be obstructed, then leading to impaired retinal perfusion. Retinal ischemia can cause the infarction of the nerve fiber layer, resulting in fluffy and white patches, called cotton wool spots (CWS). Main cause of NPDR to PDR progression is represented by an extensive retinal ischemia (9), which promotes vitreoretinal neovascularization. In fact, the retina is a high oxygen demanding tissue; under ischemia, cells release angiogenic factors, like vascular endothelial growth factor-A (VEGF-A) and placental growth factor (PlGF), which promote neovascularization. These vessels are typically fragile, fenestrated, brittle and leaky. Leaking vessels can cause vitreous hemorrhages, which are associated with gliosis and fibrovascular scar formation. Moreover, contraction of fibrous tissue can result in tractional retinal detachment and sudden loss of vision (10, 11), along with further activation of pro-fibrotic pathways. As soon as extensive vitreous hemorrhage occurs, the PDR patient is considered at high-risk of vision loss due to retinal detachment (4, 12). In case diabetic retinopathy affects the macula, the disease is also termed 'diabetic maculopathy'. Vascular leakage at the macula leads to macula swelling (diabetic macular edema, DME), which is the most common cause of blindness in diabetic patients (13, 14). DME is most prevalent during PDR, following progressive vascular and neural damage (15). Diabetic DME can be classified as 'ischemic' or 'non-ischemic', based on the involvement or preservation of the perifoveal capillary network, respectively (10). Several causative factors contribute to the pathogenesis of DME, including hypoxia and oxidative stress, upregulation of VEGF-A, alteration of the blood-retinal barrier (BRB), retinal vessel leukostasis, pericyte loss, and vascular hyperpermeability (16, 17).

THE RETINAL NEUROVASCULAR SYSTEM AS A BASIS TO UNDERSTAND DIABETIC RETINOPATHY

The retina is the innermost light-sensitive tissue of the eye, able to convert light to electrochemical signals, at first through photoreceptors, that transmit electrochemical signals to retinal neuronal circuitry (bipolar, amacrine cells). Neuroretinal electrochemical stimuli are thereafter processed and collected by retinal ganglion cells (RGCs), that transmit signals to the visual cortex by means of the optic nerve, that is constituted by RGCs axons (18, 19). The retina is characterized by a complex vascular system, whose integrity is necessary for the correct retinal function, providing nutrients and oxygen to the inner and outer retina (1). The retinal vascular system, similarly to central nervous system, is characterized by blood-retinal barrier (BRB), which maintains the right *milieu*. The BRB includes the inner and outer components. Inner BRB (iBRB) is characterized by junctions between endothelial cells (ECs) and supporting pericytes and astrocytes; while in the outer BRB (oBRB), junctions are between retinal pigmented epithelial cells (RPEs) (20, 21) (**figure 1**). Diabetes can affect both iBRB and oBRB before and after neovascular events, involving endothelial cells, pericytes (at the capillary level), vascular smooth muscle cells (arteriolar/arterial level), glia, neuronal processes, associated immune cells, and if choroid is affected, also RPEs (22).

Pericytes, endothelial cells and iBRB

Depletion of pericytes is a hallmark of DR. Pericytes wrap capillary walls and share basal lamina with endothelial cells, with which they directly interact through N-cadherin and connexin-43 hemi channels (19, 23, 24). Pericytes wrap around retinal capillaries providing structural support, modulation of endothelial cell function and homeostasis. In the inner BRB, retinal endothelial cells form the physical barriers between vascular lumen and the retina. Retinal endothelial cell-cell junctions include tight-, adherens- and gap-junction, that regulate sev-

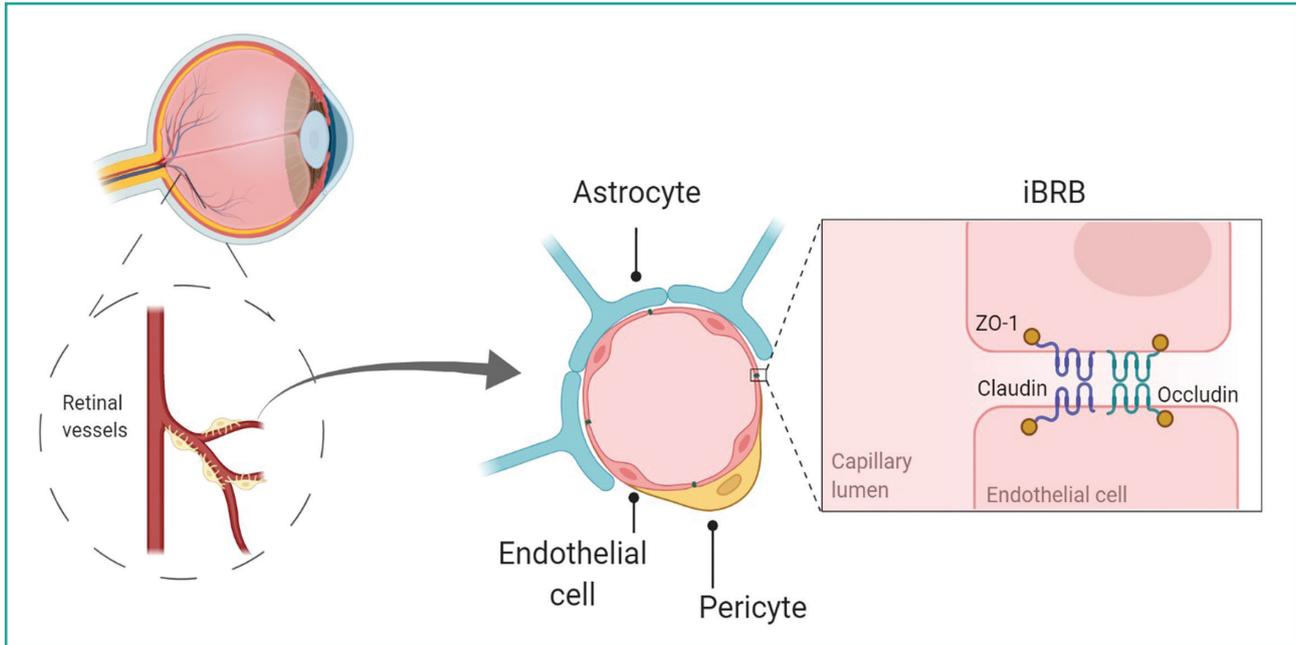


Figure 1. The neovascular unit.

In a healthy retina, vessels are structured by endothelial cells closely associated with pericytes, maintaining the function of the inner blood retina barrier (iBRB). In particular, adjacent endothelial cells are connected by tight junctions and adherens junctions.

eral cell functions such as migration, growth, protection from cell death (necrosis, apoptosis) and damage (inflammation, ischemia) (19, 25).

Retinal glia and neurons

Retinal glial cells, including Müller cells and astrocytes, provide metabolic support to neurons and play a critical role in iBRB homeostasis and integrity (26, 27). Moreover, Müller cells regulate glucose flux between the circulation and retinal neurons and have a role in providing substrates for aerobic metabolism in neurons by gluconeogenesis (28).

Immune cells

The development of new blood vessel is also supported by microglia, monocyte-derived tissue macrophages of the central nervous system. Interestingly, retinal microglia is present in the retina before development of vascularization (29). In the adult retina, ramified microglia cells were found in the inner and outer plexiform layers, and are able to produce factors that support neuronal survival. Several types of trauma

or insults lead to microglia activation, characterized by amoeboid morphology transition and production of pro-inflammatory cytokines (30). Nowadays, the improvement of the research methodologies allows us to mimic, *in vitro*, the real complexity of the ocular structures and barriers. Recently, two labs (Wisniewska-Kruk *et al.* and Fresta *et al.*) set up two different *in vitro* BRB models based on a triple co-culture of retinal cells, the first research's group used bovine cells, the second research's group used human cells. Fresta *et al.* used human retinal pericytes, astrocytes and endothelial cells to mimic the human BRB with the same cellular layer order and the same numerical ratio. This *in vitro* paradigm is useful to study and investigate the molecular mechanism related to DR, and to test new pharmacological molecules (31, 32).

PATHOPHYSIOLOGY OF DIABETIC RETINOPATHY

The pathogenesis of DR is complex and involves multiple interlinked mechanisms, in-

cluding metabolic modifications, mitochondrial dysfunction, vascular damage, apoptosis, inflammation, and oxidative stress (33-36). Several pathways have been proposed to better understand microvascular complications during DR along with sustained hyperglycemia: e.g., accumulation of advanced glycation end-products (AGEs), inflammation, activation of protein kinase C and neuronal dysfunction (19, 37). All these pathological modifications lead to increased vascular permeability and capillary depletion, resulting in macular edema and retinal neovascularization.

Hyperglycemia and Retinal Microvasculopathy

One of the earliest abnormalities observed in DR is related to retinal blood vessels, with the constriction of arteries and arterioles and blood flow anomalies (38-40). Vessel abnormalities result in a series of metabolic and biochemical alterations, like: (i) induction of activation of several PKC isoforms (e.g., PKC- α , - β , - δ and - ϵ ; in particular PKC β II isoform (41, 42); (ii) altered function of ionic channels in smooth muscle cells (BK channels) present in the retinal arteriolar vasculature (43-45). As mentioned before, retinal pericytes loss is another hallmark of the early events of DR. Several *in vitro* and *in vivo* studies report that hyperglycaemia leads to pericyte loss (34, 46, 47) or degenerated pericytes, also called "ghost cells". Therefore, pericytes loss leads to endothelial cells degeneration, microvascular destabilization and perfusion alterations with consequent ischemic events due to capillary occlusion (38, 48-50). On this regard, pericyte-like differentiation of human adipose-derived mesenchymal stem cells (hASCs) has been recently proposed as putative therapeutic tool for restoring damaged BRB (51).

Retinal inflammation

Several studies were focused on the role of inflammatory processes in early stages of DR, although, inflammatory mechanisms are still poorly understood. Chronic low-grade inflammation

has been detected in different stages of DR, both in diabetic animal models and in patients (52, 53), along with increased systemic and local expression of proinflammatory cytokines (54). In particular, microvascular endothelium, activated by these cytokines and angiogenic growth factors, expresses pro-inflammatory molecules (e.g., IL-1 β , IL-6, TNF- α , high-mobility group box-1 (HMGB1) and chemokines (MCP-1), involved in leukocyte recruitment and activation (55-57). Leukocyte-endothelium adhesion, mediated by adhesion molecules, has been implicated in leukostasis during diabetes. Sequential adhesive interactions between endothelial cells and leukocytes, are modulated by adhesion molecules (e.g., ICAM-1) present in the surface of endothelial cells, which interact with the leukocyte counter-receptor CD18 (58, 59). All these inflammatory responses may contribute to neovascularization in the retina during DR, especially under hypoxic conditions. Furthermore, increasing data suggest a crucial role of toll like receptors (TLRs) in the pathogenesis of DR; indeed, TLR4 expression is significantly increased in diabetic retinas, activating the linked inflammatory pathways (60). As regards the biochemical pathways involved in DR, expression of inflammatory cytokines might be mediated by activation of mitogen-activated protein kinases (MAPKs) (61), as well as ERKs, normally involved in several cellular processes (62). ERK pathway can influence NF- κ B activation, by the regulation of NF- κ B-dependent genes expression, e.g. inducible nitric oxide synthase (iNOS), cyclooxygenase-2 (COX-2) and tumor necrosis factor-alpha (TNF- α) (63).

Retinal hypoxia

The retina is one of the most oxygen and glucose demanding tissue (64). Retinal hypoxia represents an important causative factor for DR development, and plays a central role in progression of NPDR to PDR, due to the release of some soluble mediators such as cytokines, chemokine and growth factors, which promote the growth of extraretinal neovascularization (65). Ocular ischemic events are

considered crucial for promotion of vascular abnormalities, due to endothelial cells adaptation to stress, which upregulate several genes, like VEGF-A (66). Furthermore, VEGF-A and other hypoxia-regulated growth factors, are controlled by hypoxia-inducible factor (HIF) (67). HIF is a heterodimer, HIF-1 α (inducible subunit) and HIF-1 β (constitutively expressed). Oxygen deprivation, induces HIF-1 α to translocate into the nucleus and to bind the hypoxia-response elements (HREs) in DNA, leading to expression of inflammatory and pro-angiogenic genes, promoting inflammation and angiogenesis, respectively (68, 69).

Retinal angiogenesis

Angiogenesis is a crucial mechanism in physiological vascular development and during

pathological conditions. Angiogenesis is related to ECs that, stimulated by some angiogenic factors, generate new blood vessels (70). Indeed, this process is characterized by the angiogenic growth factors, which activate the receptors present on resident ECs; then, endothelial cells begin to release specific enzymes such as matrix metalloproteinases (MMPs) which degrade the basement membrane, leading ECs to leave the original vessel wall. After that, endothelial cells start to proliferate into the surrounding matrix, thanks to the adhesion molecules (**figure 2**).

The main regulators of angiogenesis are the vascular endothelial growth factors (VEGF-A, VEGF-B, VEGF-C, and VEGF-D) and the placental growth factor (PlGF) (71-76). VEGFs can bind to three tyrosine kinase receptors:

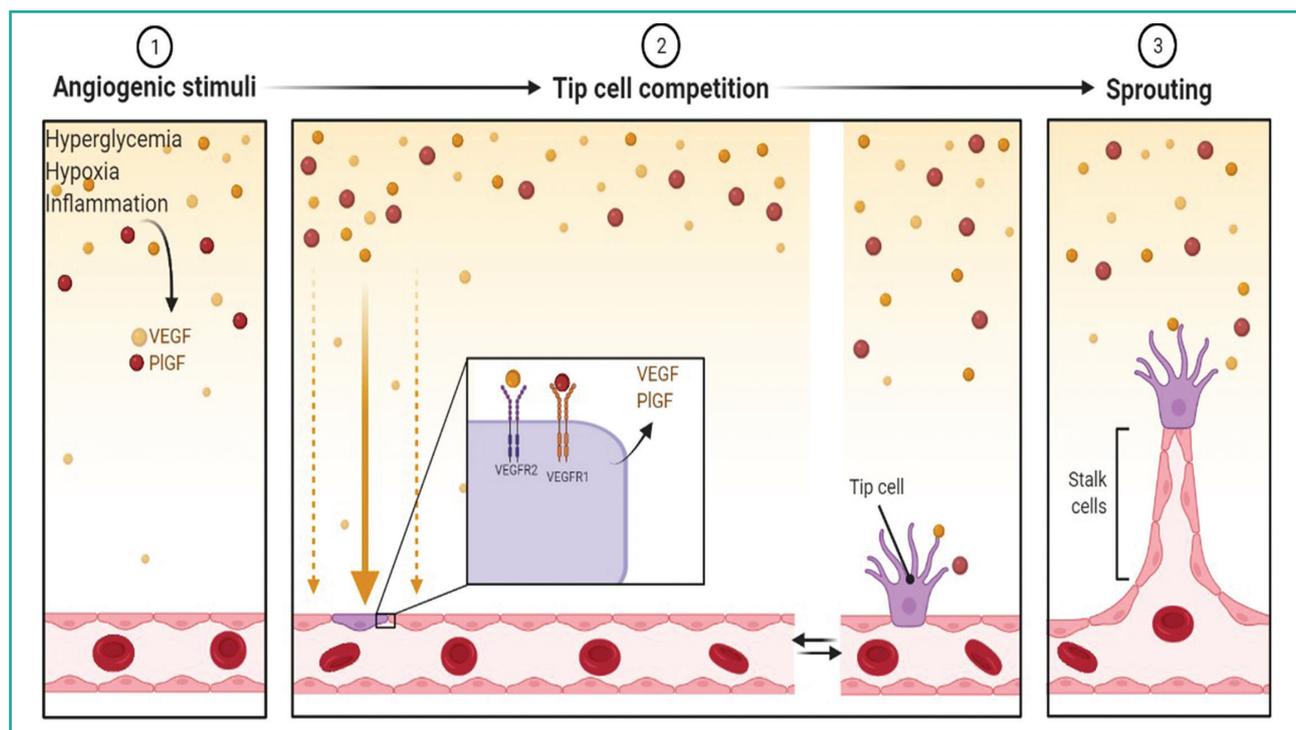


Figure 2. Retinal angiogenesis in diabetic retinopathy.

Angiogenic factors (*i.e.*, VEGF-A and PlGF) stimulate angiogenesis in tissues. VEGF-A/PlGF bind to VEGFR-2/VEGFR-1 on the surface of endothelial cells (ECs), triggering competition between neighboring cells as they differentiate. In normal intact retinal vessels, blood flow is regular and the vascular configuration is stable. However, in diabetic retinopathy, this process is strongly exacerbated by several phenomena, such as hyperglycemia, hypoxia and inflammation. The expression of angiogenic factors is increased and the growth of new blood vessels is uncontrolled. The neovascularization is typical of the later stages of diabetic retinopathy, with the formation of new unstable vessels. This leads to vascular damage, loss of endothelial tight junctions, pericytes detachment and basement membrane thickening (iBRB breakdown).

VEGFR-1 (Flt-1), -2 (KDR), and -3 (Flt-4) (77). VEGFR-1 (78) and VEGFR-2 (79–81) are the main receptors involved in angiogenesis. VEGFR-2 (also known as Flk1) is expressed on endothelial cells. Binding to VEGFR-1 (also known as Flt1) leads to the activation of quiescent endothelial cells and promote vascular permeability (82-85). VEGF-A is significantly increased in ocular tissues from patients with diabetes (86). All the mechanisms linked to the progression of DR, are responsible of the overproduction of VEGF-A, including hypoxic events. Besides stimulation of endothelial cell growth, VEGF-A can also promote the disassembly of junctions between endothelial cells, leading to vascular permeability (BRB breakdown).

Fibrosis

Angiogenesis and subsequent fibrotic events occur with progression of PDR. Fibrosis can cause the formation of fibrovascular epiretinal membranes, which lead to retinal complications such as tractional retinal detachment and, at last, vision loss (87-89). Fibrosis is a complex reparative process that is activated to restore damaged tissue, by means of remodelling extracellular matrix (ECM). Cell proliferation, ECM deposition and neovascularization are key mechanisms during PDR, usually stimulated by pathological conditions like hypoxia or inflammation, promoting formation of fibrotic tissue (90, 91). Along with microglia and astrocytes, Muller cells in response to retinal injury, participate to fibrotic events, through production of inflammatory and angiogenic mediators (92, 93). Fibrosis can also be promoted by retinal hypoxia, leading to a consequent overproduction of VEGF-A (94-96). Several growth factors play a role in fibrosis, such as transforming growth factor- β (TGF- β), platelet-derived growth factor (PDGF), and the pro-fibrotic connective tissue growth factor (CTGF) (97-99). Precisely, increased levels of CTGF were found in the vitreous of patient with PDR (100, 101) and it has been supposed that CTGF could be a downstream

mediator of TGF- β , the main regulator of pro-fibrotic effects.

PHARMACOLOGICAL TREATMENT OF DR

Currently, only PDR can be pharmacologically treated, and no approved treatments are available for NPDR. As mentioned above, the hallmarks of this disease are the abnormal vessel growth in retinal area, up-regulation of inflammatory factors, and the breakdown of the blood-retinal barrier (**figure 3 A**). Clinical history of PDR has been revolutionized with anti-angiogenic treatments, that are invasive and expansive. Along with the anti-VEGF agents (34), anti-inflammatory drugs are also used (102). Steroids are potent drugs to quench inflammation and reduce edema, fibrin deposition, capillary hyperpermeability and phagocytic migration typical of the inflammatory response (103-105). Furthermore, they also counteract the action of VEGF-A (106). Three corticosteroids are actually approved to handle diabetic macular edema (DME): dexamethasone (DEX), fluocinolone acetonide (FA) and triamcinolone acetonide (TA). The limitation of these drugs is related to the side effects such as cataract and rise in intraocular pressure (102).

Anti-VEGF therapy

The anti-VEGF therapies have revolutionized the treatment of DR. These medications, such as ranibizumab (Lucentis, Genentech) and aflibercept (Eylea, Regeneron), called vascular endothelial growth factor inhibitors (anti-VEGF), have a consolidate history in terms of efficacy and safety for the treatment of DME. Ranibizumab is a 48 kDa antigen-binding fragment (Fab) of a humanized monoclonal antibody with high affinity for VEGF-A (**figure 3 B**) (107); it binds with high affinity all the VEGF-A isoforms (such as VEGF-A₁₆₅, VEGF-A₁₁₀ and VEGF-A₁₂₁) reducing the activation of VEGFR-1 and VEGFR-2 receptors. The small size of this fragment enhances its diffusion from the vitreous to the retina and the choroid, improving

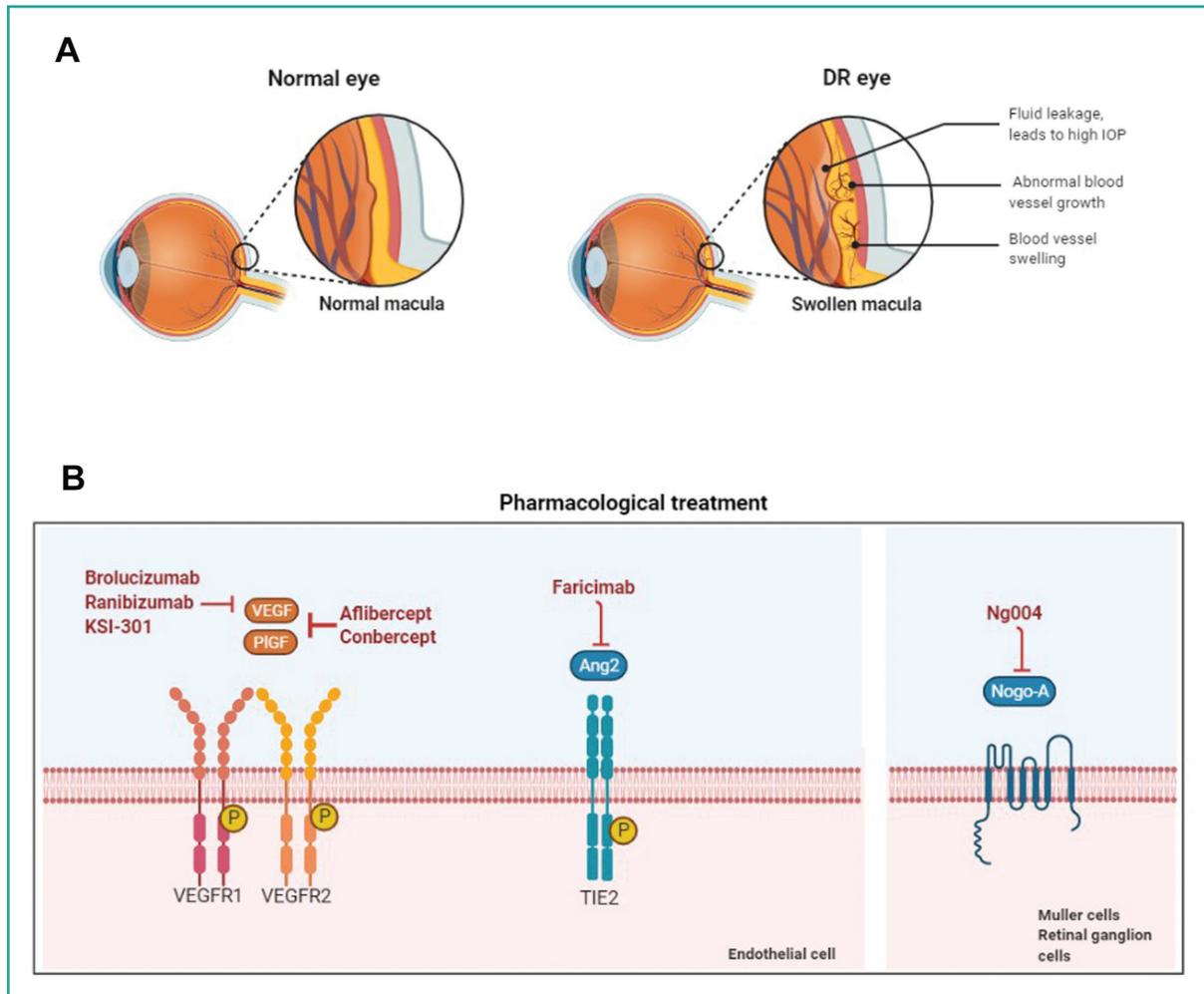


Figure 3. Diabetic retinopathy clinical hallmarks and treatments. **(A)** Diabetic retinopathy is the leading cause of vision loss in diabetic patients and is characterized by abnormal vessel growth in retinal area, inflammation, breakdown of the blood-retinal barrier and fluid accumulation. **(B)** Therapeutic strategies targeting different signalling pathways involved in the pathogenesis of proliferative DR and diabetic macular edema.

the pharmacokinetic profile, compared to bevacizumab (108).

Aflibercept has been approved by Food and Drug Administration (FDA) in 2011 for the treatment of age-related macular degeneration (AMD), for impaired vision due to secondary macular edema, caused by retinal vein occlusion (Branch RVO or central RVO) and for the treatment of visual impairment due to myopic choroidal neovascularization (CNV). Recently, aflibercept was also approved for the treatment of diabetic macular edema. Aflibercept (VEGF-trap) is a fusion protein (115 kDa) bearing two binding domains of VEGF receptors (**figure 3**

B) (109). Moreover, aflibercept's binding affinity to VEGF-A₁₆₅ is almost 100-fold greater than ranibizumab and bevacizumab (110-112), and is the only anti-VEGF agents that binds PlGF, although with lower affinity (38.9 nM dissociation constant – KD), compared to VEGF-A (0.49 nM dissociation constant – KD) (113).

Conbercept (Lumitin) is a 141 kDa recombinant fusion protein composed of the second Ig domain of VEGFR-1 and the third and fourth Ig domains of VEGFR-2, fused to the constant region (Fc) of human IgG1 (**figure 3 B**). Considering the increasing need for less frequent intravitreal injections of anti-VEGF, conbercept

has designed to improve dose regimens and compliance. Similarly to aflibercept, conbercept has multiple targets (114).

In 2021 FDA approved the faricimab (Roche) with the following indications: wet AMD and DME. This antibody targets two different pathways involved in progression of these retinal diseases: angiopoietin-2 (Ang-2) and vascular endothelial growth factor-A (VEGF-A) (**figure 3 B**). Faricimab showed positive results across four phase III studies in AMD and DME. Faricimab clinical trials proved a non-inferiority efficacy evidence, compared to aflibercept (115, 116).

Brolucizumab (Beovu, Novartis), recently approved in US and EU, is a humanized single-chain antibody fragment (scFv) targeting three major isoforms of VEGF-A (e.g., VEGF₁₁₀, VEGF₁₂₁, and VEGF₁₆₅) (**figure 3 B**). Compared with other VEGF-A inhibitors, brolucizumab is smaller (26 kDa). In 2021 Novartis announced the positive results of the Phase III KESTREL study. This study assessed safety and efficacy of 6 mg brolucizumab in patients with DME.

Although most of patients show beneficial effect by the approved anti-VEGF agents, a significant percentage of people are poor responders. To overcome this unmet medical need, NovaGo Therapeutics is developing a first-in-class fully human antibody therapy with a novel mechanism of action (NG004) targeting the protein Nogo-A. This latter is endogenously expressed by Müller cells and retinal ganglion cells, and it represents one of the most up-regulated protein during DR (117, 118). It has been demonstrated that the block of this protein could lead to the reduction of angiogenesis and inflammation, as already demonstrated in an *in vivo* model of retinal injury (excitotoxicity-induced neuroinflammation) (119).

As regards the safety profile of intravitreal anti-VEGF agents some adverse reactions such as endophthalmitis, intraocular inflammation, intraocular pressure elevation and ocular hemorrhage are sometimes associated with the treatment (120-122). Besides that, VEGF protein has a physiological role in the retina, so

the prolonged period of treatment with these compounds could be deleterious (123). Moreover, these agents have a short half-life, and the widely treatment schedule is the treat-and-extend regimen with several injections for several months. For these reasons new agents and innovative delivery systems are under investigation.

NOVEL MOLECULAR TARGETS

Placental Growth Factor (PIGF)

PIGF has been implicated in pathological angiogenesis, especially in retinal disorders, although its function is less well understood (85), compared to VEGF-A. Oppositely to VEGF-A, PIGF is not required during physiological angiogenesis but plays a role only during pathological conditions (82, 83, 112, 124-128). Secreted PIGF specifically acts through VEGFR-1. Furthermore, it has been showed that VEGF-A and PIGF can form heterodimers (129) which can bind both VEGFR-1 and VEGFR-2, stimulating endothelial cells migration and vasorelaxation via the nitric oxide pathway (130, 131). Moreover, it has been found that VEGF/PIGF heterodimer can lead to activation of a positive feedback and overproduction of VEGF-A, which binds also the VEGFR-2. Therefore, PIGF may stimulate angiogenesis directly through VEGFR-1 but also indirectly through VEGFR-2 (83, 128). PIGF acts also through neuropilin receptor 1 (NRP1) (124, 127, 132), that is expressed in angiogenic vessels (133, 134). As well as VEGF-A, PIGF is expressed by endothelial cells in hypoxic environment (135-137). The PIGF overexpression is driven by HIF-1 α , which is able to recognize a hypoxia responsive element (HRE) located in the second intron of PIGF gene (136). One interesting recent evidence demonstrated that aflibercept and a specific anti-PIGF antibody exert anti-inflammatory effects in the diabetic retina. Specifically, aflibercept and anti-PIGF antibody protected retinal endothelial cells (HRECs) and primary mouse retinal pigmented epithelial

cells (mRPEs) from cell damage induced by high glucose levels, blocking the activation of the ERK pathway with the subsequent suppression of TNF- α release (113).

miRNAs

There is an increasing interest on microRNAs as putative biomarkers for the progression of DR (138). Platania *et al.*, demonstrated that small set of miRNAs were dysregulated in serum and retina of diabetic mice. These miRNAs were also dysregulated in serum of patients with diabetic retinopathy. In the *in-vivo* study, these miRNAs modulated not only VEGF-A expression (up-regulation) but also the neurotrophic factor BDNF (down-regulated) (139). Moreover, Santovito *et al.*, reported that DR is associated with higher circulating levels of miR-25-3p and miR-320b and lower levels of miR-495-3p, in patients with type 2 diabetes and diabetic retinopathy (140). Interestingly, it has been demonstrated a specific association between miRNAs expression and hypoxic microenvironment; in fact, retinal

hypoxia led to the upregulation of six miRNAs (miR-20a-5p, miR-20b-5p, miR-27a-3p, miR-27b-3p, miR-206-3p, miR-381-3p) in human retinal endothelial cells. These miRNAs, are capable to interfere with the expression of genes belonging to the TGF- β pathway at post-transcriptional level. In fact, the dysregulation of these miRNAs has driven and promoted angiogenesis and fibrosis, through the modulation of VEGF-A, TGF- β and HIF-1 α , in retinal endothelial cells (135, 141-144). Moreover, Shao *et al.* identified miR-136 and miR-374 dysregulation as hallmark of proliferative DR (145). The putative involvement of miRNAs in the pathogenesis of DR is also linked to the direct activation of the inflammatory pathway through the TLR-4, as well demonstrated in several *in vitro* and *in vivo* models of DR; in fact, different miRNAs are associated with the regulation of TLR-4 expression during diabetic retinopathy (146-148). Currently, evidence about post-transcriptional regulation of PlGF expression by miRNAs has not been retrieved. A high-throughput screening of miR-

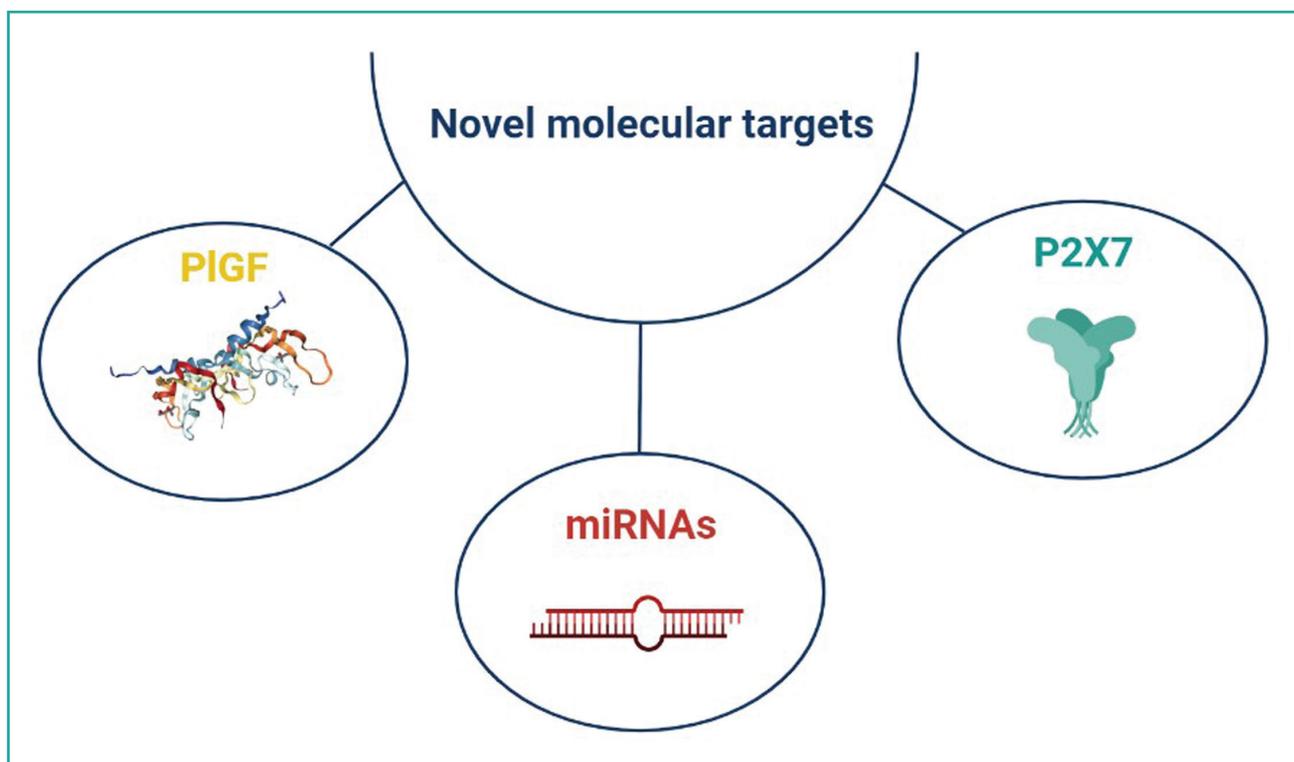


Figure 4. Novel molecular targets for the treatment of diabetic retinopathy.

NAs potentially related to PIGF would address this issue.

P2X7 receptor

In the last decade an important link between DR and purinergic receptor has been demonstrated, considering P2X7 receptor as a putative pharmacological target in this retinal disease (149-151). P2X7R is a member of the family of purinoceptors, ligand-gated membrane ion channels activated by extracellular ATP. This receptor is widely distributed in all retinal layers and also in retinal microvasculature. P2X7R stimulation promotes a wide range of cellular responses, ranging from proliferation to cell death, from cytokines release to reactive oxygen species (ROS) production. The early up-regulation and activation of P2X7R has been related to several types of retinal diseases, and its antagonism revealed benefits against inflammation, oxidative stress and angiogenesis, both *in vitro* and *in vivo* studies (152-156). In particular, the selective antagonist of this receptor (JNJ47965567) has shown anti-inflammatory effect, through the decreased activation of inflammasome and IL-1 β production, in several pathological conditions. Moreover, P2X7R inhibition up-regulated the expression of junction proteins in the iBRB, which is compromised in early DR (156, 157). Furthermore, it was found a significant activation of P2X7R during retinal hypoxia, and the P2X7R blockade, by selective P2X7R antagonists A740003 and AZ10606120, inhibited the HIF-1 α and VEGF-A retinal overexpression (151, 158).

CONCLUSIONS

Increasing evidence suggest that retinal neurodegeneration and inflammation are implicated in the pathogenesis of diabetic retinopathy. Several recent studies were carried out to explore new pharmacological targets, potentially able to counteract retinal neurodegeneration and inflammation. The present review summarizes new hints and puzzle pieces about the etiopathogenesis of DR, addressing several hypotheses and trying to identify and vali-

date novel and promising pathways implicated in this pathology. Currently, the first-line pharmacological therapy for PDR and DME is represented by intravitreal injection of anti-VEGF agents and corticosteroids, respectively. However, the current proliferative diabetic retinopathy pharmacotherapy is characterized by frequent, invasive and expensive treatments, that have a significant impact on health system. There are several unmet medical needs in the management of DR that stimulate the pharmacological research to develop novel pharmacological targets and drugs to counteract the early phases of diabetic retinopathy.

ACKNOWLEDGEMENTS

This work was supported by the Italian Ministry of Economic Development (MISE) PON-Innovative PhD Program. The author would like to thank the Pharmacology section of the University of Catania, in particular Prof. Claudio Bucolo, Dr. Chiara Bianca Maria Platania and Dr. Federica Conti for reviewing the manuscript.

ETHICS

Fundings

Ministry of University and Research (MUR) PON-Innovative PhD Program #E62C17000000006.

Conflict of interests

The authors declare that they have no conflict of interests.

Authors' contribution

All the authors contributed equally to conception, data collection, analysis and writing of this paper.

Availability of data and materials

N/A

Ethical approval

N/A

REFERENCES

1. Cheloni R, Gandolfi SA, Signorelli C, Odone A. Global prevalence of diabetic retinopathy: Protocol for a systematic review and meta-analysis. *BMJ Open*. 2019;9(3):e022188. doi: 10.1136/bmjopen-2018-022188.
2. Ruta LA, Magliano DJ, LeMesurier R, Taylor HR, Zimmet PZ, Shaw JE. Prevalence of diabetic retinopathy in type 2 diabetes in developing and developed countries. *Diabet Med*. 2013;30(4):387-98. doi: 10.1111/dme.12119.
3. Zhang X, Saaddine JB, Chou CF, Cotch MF, Cheng YJ, Geiss LS, et al. Prevalence of diabetic retinopathy in the United States, 2005-2008. *JAMA* 2010;304(6):649-56. doi: 10.1001/jama.2010.1111.
4. American Academy of Ophthalmology. Preferred Practice Pattern® Guidelines: Diabetic Retinopathy. *Am Acad Ophthalmol*. 2016.
5. Kashim RM, Newton P, Ojo O. Diabetic retinopathy screening: A systematic review on patients' non-attendance. *Int J Environ Res Publ Health*. 2018;15(1):157. doi: 10.3390/ijerph15010157.
6. Maric-Bilkan C. Sex differences in micro- and macro-vascular complications of diabetes mellitus. *Clin Sci (Lond)*. 2017;131(9):833-46. doi: 10.1042/CS20160998.
7. Shukla U V, Tripathy K. Diabetic Retinopathy. *StatPearls*. Treasure Island (FL) 2022.
8. Tarr JM, Kaul K, Chopra M, Kohner EM, Chibber R. Pathophysiology of Diabetic Retinopathy. *ISRN Ophthalmol*. 2013;2013:343560. doi: 10.1155/2013/343560.
9. Duh EJ, Sun JK, Stitt AW. Diabetic retinopathy: current understanding, mechanisms, and treatment strategies. *JCI insight*. 2017;2(14):e93751. doi: 10.1172/jci.insight.93751.
10. Stitt AW, Curtis TM, Chen M, Medina RJ, McKay GJ, Jenkins A, et al. The progress in understanding and treatment of diabetic retinopathy. *Prog Retin Eye Res*. 2016;51:156-86. doi: 10.1016/j.preteyeres.2015.08.001.
11. Lechner J, O'Leary OE, Stitt AW. The pathology associated with diabetic retinopathy. *Vision Res*. 2017;139:7-14. doi: 10.1016/j.visres.2017.04.003.
12. Poretzky L. Principles of diabetes mellitus. *Principles of Diabetes Mellitus*. 2010.
13. Wu L. Classification of diabetic retinopathy and diabetic macular edema. *World J Diabetes*. 2013; 4(6):290-4. doi: 10.4239/wjd.v4.i6.290.
14. Wilkinson CP, Ferris FL, Klein RE, Lee PP, Agardh CD, Davis M, et al. Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales. *Ophthalmology*. 2003;110(9):1677-82. doi: 10.1016/S0161-6420(03)00475-5.
15. Antonetti DA, Klein R, Gardner TW. Diabetic retinopathy. *N Engl J Med*. 2012;366(13):1227-39. doi: 10.1056/NEJMra1005073.
16. Chung YR, Kim YH, Ha SJ, Byeon HE, Cho CH, Kim JH, et al. Role of Inflammation in Classification of Diabetic Macular Edema by Optical Coherence Tomography. *J Diabetes Res* 2019;8164250. doi: 10.1155/2019/8164250.
17. Ascaso FJ, Huerva V, Grzybowski A. The role of inflammation in the pathogenesis of macular edema secondary to retinal vascular diseases. *Mediators Inflamm*. 2014;2014:432685. doi: 10.1155/2014/432685.
18. Hoon M, Okawa H, Della Santina L, Wong ROL. Functional architecture of the retina: Development and disease. *Prog Retin Eye Res*. 2014;42:44-84. doi: 10.1016/j.preteyeres.2014.06.003.
19. Shin ES, Sorenson CM, Sheibani N. Diabetes and Retinal Vascular Dysfunction. *J Ophthalmic Vis Res*. 2014;9(3):362-73. doi: 10.4103/2008-322X.143378.
20. Alizadeh E, Mammadzada P, André H. The different facades of retinal and chorioidal endothelial cells in response to hy-

- poxia. *Int J Mol Sci.* 2018;19(12):3846. doi: 10.3390/ijms19123846.
21. Abbott NJ, Patabendige AAK, Dolman DEM, Yusof SR, Begley DJ. Structure and function of the blood-brain barrier. *Neurobiol Dis.* 2010;37(1):13-25. doi: 10.1016/j.nbd.2009.07.030.
 22. Klaassen I, Van Noorden CJF, Schlingemann RO. Molecular basis of the inner blood-retinal barrier and its breakdown in diabetic macular edema and other pathological conditions. *Prog Retin Eye Res.* 2013;34:19-48. doi: 10.1016/j.preteyeres.2013.02.001.
 23. Armulik A, Abramsson A, Betsholtz C. Endothelial/pericyte interactions. *Circ Res.* 2005;97(6):512-23. doi: 10.1161/01.RES.0000182903.16652.d7.
 24. Bobbie MW, Roy S, Trudeau K, Munger SJ, Simon AM, Roy S. Reduced connexin 43 expression and its effect on the development of vascular lesions in retinas of diabetic mice. *Invest Ophthalmol Vis Sci.* 2010;51(7):3758-63. doi: 10.1167/iovs.09-4489.
 25. Linder S. The matrix corroded: podosomes and invadopodia in extracellular matrix degradation. *Trends Cell Biol.* 2007;17(3):107-17. doi: 10.1016/j.tcb.2007.01.002.
 26. Bringmann A, Pannicke T, Grosche J, Francke M, Wiedemann P, Skatchkov SN, et al. Müller cells in the healthy and diseased retina. *Prog Retin Eye Res.* 2006;25(4):397-424. doi: 10.1016/j.preteyeres.2006.05.003.
 27. Bringmann A, Wiedemann P. Müller glial cells in retinal disease. *Ophthalmologica.* 2012;227(1):1-19. doi: 10.1159/000328979.
 28. Newman E, Reichenbach A. The Muller cell: A functional element of the retina. *Trends Neurosci.* 1996;19(8):307-12. doi: 10.1016/0166-2236(96)10040-0.
 29. Dejda A, Mawambo G, Cerani A, Miloudi K, Shao Z, Daudelin JF, et al. Neuropilin-1 mediates myeloid cell chemoattraction and influences retinal neuroimmune crosstalk. *J Clin Invest.* 2014;124(11):4807-22. doi: 10.1172/JCI76492.
 30. Langmann T. Microglia activation in retinal degeneration. *J Leukoc Biol.* 2007;81(6):1345-51. doi: 10.1189/jlb.0207114.
 31. Fresta CG, Fidilio A, Caruso G, Caraci F, Giblin FJ, Leggio GM, et al. A new human blood-retinal barrier model based on endothelial cells, pericytes, and astrocytes. *Int J Mol Sci.* 2021;162. doi: 10.3390/ijms21051636.
 32. Wisniewska-Kruk J, Hoeben KA, Vogels IMC, Gaillard PJ, van Noorden CJF, Schlingemann RO, et al. A novel co-culture model of the blood-retinal barrier based on primary retinal endothelial cells, pericytes and astrocytes. *Exp Eye Res.* 2012;96(1):181-90. doi: 10.1016/j.exer.2011.12.003.
 33. Kern TS, Antonetti DA, Smith LEH. Pathophysiology of Diabetic Retinopathy: Contribution and Limitations of Laboratory Research. *Ophthalmic Res.* 2019;62(4):196-202. doi: 10.1159/000500026.
 34. Wang W, Lo ACY. Diabetic retinopathy: Pathophysiology and treatments. *Int J Mol Sci.* 2018;19(6):1816. doi: 10.3390/ijms19061816.
 35. Wu MY, Yiang GT, Lai TT, Li CJ. The oxidative stress and mitochondrial dysfunction during the pathogenesis of diabetic retinopathy. *Oxid Med Cell Longev* 2018;3420187. doi: 10.1155/2018/3420187.
 36. Heng LZ, Comyn O, Peto T, Tadros C, Ng E, Sivaprasad S, et al. Diabetic retinopathy: Pathogenesis, clinical grading, management and future developments. *Diabet Med.* 2013; 30(6):640-50. doi: 10.1111/dme.12089.
 37. Bhagat N, Grigorian RA, Tutela A, Zarbin MA. Diabetic Macular Edema: Pathogenesis and Treatment. *Surv Ophthalmol.* 2009;54(1):1-32. doi: 10.1016/j.survophthal.2008.10.001.

38. Durham JT, Herman IM. Microvascular modifications in diabetic retinopathy. *Curr Diab Rep.* 2011;11(4):253-64. doi: 10.1007/s11892-011-0204-0.
39. Zhu Q, Xu X, Xia X, Gu Q, Ho PCP. Role of protein kinase C on the alteration of retinal endothelin-1 in streptozotocin-induced diabetic rats. *Exp Eye Res.* 2005;81(2):200-6. doi: 10.1016/j.exer.2005.01.025.
40. Wong TY, Klein R, Richey Sharrett A, Schmidt MI, Pankow JS, Couper DJ, et al. Retinal arteriolar narrowing and risk of diabetes mellitus in middle-aged persons. *JAMA.* 2002;287(19):2528-33. doi: 10.1001/jama.287.19.2528.
41. Kim JH, Kim JH, Jun HO, Yu YS, Kim KW. Inhibition of protein kinase C δ attenuates blood-retinal barrier breakdown in diabetic retinopathy. *Am J Pathol.* 2010;176(3):1517-24. doi: 10.2353/ajpath.2010.090398.
42. Amadio M, Bucolo C, Leggio GM, Drago F, Govoni S, Pascale A. The PKC β /HuR/VEGF pathway in diabetic retinopathy. *Biochem Pharmacol.* 2010; 80(8):1230-7. doi: 10.1016/j.bcp.2010.06.033.
43. McGahon MK, Dash DP, Arora A, Wall N, Dawicki J, Simpson DA, et al. Diabetes downregulates large-conductance Ca²⁺-activated potassium β 1 channel subunit in retinal arteriolar smooth muscle. *Circ Res.* 2007;100(5):703-11. doi: 10.1161/01.RES.0000260182.36481.c9.
44. Curtis TM, Gardiner TA, Stitt AW. Microvascular lesions of diabetic retinopathy: Clues towards understanding pathogenesis? *Eye.* 2009;23(7):1496-508. doi: 10.1038/eye.2009.108.
45. Barot M, Gokulgandhi MR, Patel S, Mitra AK. Microvascular complications and diabetic retinopathy: Recent advances and future implications. *Future Med Chem.* 2013;5(3):301-14. doi: 10.4155/fmc.12.206.
46. Naruse K, Nakamura J, Hamada Y, Nakayama M, Chaya S, Komori T, et al. Aldose reductase inhibition prevents glucose-induced apoptosis in cultured bovine retinal microvascular pericytes. *Exp Eye Res.* 2000;71(3):309-15. doi: 10.1006/exer.2000.0882.
47. Romeo G, Liu WH, Asnaghi V, Kern TS, Lorenzi M. Activation of nuclear factor- κ B induced by diabetes and high glucose regulates a proapoptotic program in retinal pericytes. *Diabetes.* 2002;51(7):2241-8. doi: 10.2337/diabetes.51.7.2241.
48. Benjamin LE, Hemo I, Keshet E. A plasticity window for blood vessel remodeling is defined by pericyte coverage of the preformed endothelial network and is regulated by PDGF-B and VEGF. *Development.* 1998;125(9):1591-8. doi: 10.1242/dev.125.9.1591.
49. Orlidge A, D'Amore PA. Inhibition of capillary endothelial cell growth by pericytes and smooth muscle cells. *J Cell Biol.* 1987; 105(3):1455-62. doi: 10.1083/jcb.105.3.1455.
50. Beltramo E, Porta M. Pericyte Loss in Diabetic Retinopathy: Mechanisms and Consequences. *Curr Med Chem.* 2013; 20(26):3218-25. doi: 10.2174/09298673113209990022.
51. Mannino G, Gennuso F, Giurdanella G, Conti F, Drago F, Salomone S, et al. Pericyte-like differentiation of human adipose-derived mesenchymal stem cells: An in vitro study. *World J Stem Cells.* 2020;26;12(10):1152-70.
52. Miyamoto K, Khosrof S, Bursell SE, Rohan R, Murata T, Clermont AC, et al. Prevention of leukostasis and vascular leakage in streptozotocin-induced diabetic retinopathy via intercellular adhesion molecule-1 inhibition. *Proc Nat Acad Sci USA.* 1999; 96(19):10836-41. doi:10.1073/pnas.96.19.10836.
53. Yuuki T, Kanda T, Kimura Y, Kotajima N, Tamura J, Kobayashi I, et al. Inflammatory cytokines in vitreous fluid and serum of patients with diabetic vitreoretinopathy. *J Diabetes Complications.* 2001;15(5):257-9. doi: 10.1016/s1056-8727(01)00155-6.

54. Tang J, Kern TS. Inflammation in diabetic retinopathy. *Prog Retin Eye Res.* 2011;30(5):343-58. doi: 10.1016/j.preteyeres.2011.05.002.
55. Newton K, Dixit VM. Signaling in innate immunity and inflammation. *Cold Spring Harb Perspect Biol.* 2012;4(3):a006049. doi: 10.1101/cshperspect.a006049.
56. Speyer CL, Ward PA. Role of endothelial chemokines and their receptors during inflammation. *J Invest Surg.* 2011;24(1):18-27. doi: 10.3109/08941939.2010.521232.
57. Sprague AH, Khalil RA. Inflammatory cytokines in vascular dysfunction and vascular disease. *Biochem Pharmacol.* 2009;78(6):539-52. doi:10.1016/j.bcp.2009.04.029.
58. Jousen AM, Poulaki V, Le ML, Koizumi K, Esser C, Janicki H, et al. A central role for inflammation in the pathogenesis of diabetic retinopathy. *FASEB J.* 2004;18(12):1450-2. doi: 10.1096/fj.03-1476fje.
59. Al-Kharashi AS. Role of oxidative stress, inflammation, hypoxia and angiogenesis in the development of diabetic retinopathy. *Saudi J Ophthalmol.* 2018;32(4):318-23. doi:10.1016/j.sjopt.2018.05.002.
60. Bayan N, Yazdanpanah N, Rezaei N. Role of toll-like receptor 4 in diabetic retinopathy. *Pharmacol Res.* 2022;175:105960. doi:10.1016/j.phrs.2021.105960.
61. Son Y, Cheong Y-K, Kim N-H, Chung H-T, Kang DG, Pae H-O. Mitogen-Activated Protein Kinases and Reactive Oxygen Species: How Can ROS Activate MAPK Pathways? *J Signal Transduct.* 2011;2011:792639. doi: 10.1155/2011/792639.
62. Cargnello M, Roux PP. Activation and Function of the MAPKs and Their Substrates, the MAPK-Activated Protein Kinases. *Microbiol Mol Biol Rev.* 2011;75(1):50-83. doi: 10.1128/MMBR.00031-10.
63. Jiang B, Xu S, Hou X, Pimentel DR, Brecher P, Cohen RA. Temporal Control of NF- κ B Activation by ERK Differentially Regulates Interleukin-1 β -induced Gene Expression. *J Biol Chem.* 2004;279(2):1323-9. doi: 10.1074/jbc.M307521200.
64. Ames A. Energy requirements of CNS cells as related to their function and to their vulnerability to ischemia: A commentary based on studies on retina. *Can J Physiol Pharmacol.* 1992;70 Suppl:S158-64. doi: 10.1139/y92-257.
65. Dell'Omo R, Semeraro F, Bamonte G, Cifariello F, Romano MR, Costagliola C. Vitreous mediators in retinal hypoxic diseases. *Mediators Inflamm.* 2013. doi: 10.1155/2013/935301.
66. Paternotte E, Gaucher C, Labrude P, Stoltz JF, Menu P. Review: Behaviour of endothelial cells faced with hypoxia. *Bio-Med Mater Eng.* 2008;18(4-5):295-9. PMID: 19065037.
67. Wang GL, Semenza GL. General involvement of hypoxia-inducible factor 1 in transcriptional response to hypoxia. *Proc Nat Acad Sci USA.* 1993;90(9):4304-8. doi: 10.1073/pnas.90.9.4304.
68. Peet DJ, Kittipassorn T, Wood JP, Chidlow G, Casson RJ. HIF signalling: The eyes have it. *Exp Cell Res.* 2017;356(2):136-40. doi: 10.1016/j.yexcr.2017.03.030.
69. Vadlapatla R, Vadlapudi A, Mitra A. Hypoxia-Inducible Factor-1 (HIF-1): A Potential Target for Intervention in Ocular Neovascular Diseases. *Curr Drug Targets.* 2013;14(8):919-35. doi: 10.2174/13894501113149990015.
70. Potente M, Gerhardt H, Carmeliet P. Basic and therapeutic aspects of angiogenesis. *Cell.* 2011;146(6):873-87. doi: 10.1016/j.cell.2011.08.039.
71. Ferrara N. Role of vascular endothelial growth factor in regulation of physiological angiogenesis. *Am J Physiol - Cell Physiol.* 2001;280(6):C1358-66. doi: 10.1152/ajpcell.2001.280.6.C1358.
72. Errico M, Riccioni T, Iyer S, Pisano C, Acharya KR, Persico MG, et al. Identification of placenta growth factor determinants for binding and activation of Flt-1 recep-

- tor. *J Biol Chem.* 2004;279(42):43929-39. doi: 10.1074/jbc.M401418200.
73. Dewerchin M, Carmeliet P. Placental growth factor in cancer. *Expert Opin Ther Targets.* 2014;18(11):1339-54. doi: 10.1517/14728222.2014.948420.
 74. Hiratsuka S, Maru Y, Okada A, Seiki M, Noda T, Shibuya M. Involvement of Flt-1 tyrosine kinase (vascular endothelial growth factor receptor-1) in pathological angiogenesis. *Cancer Res.* 2001;61(3):1207-13. PMID: 11221852.
 75. Robinson CJ, Stringer SE. The splice variants of vascular endothelial growth factor (VEGF) and their receptors. *J Cell Sci.* 2001;114(Pt 5):853-65. doi: 10.1242/jcs.114.5.853.
 76. Fontanella C, Ongaro E, Bolzonello S, Guardascione M, Fasola G, Aprile G. Clinical advances in the development of novel VEGFR2 inhibitors. *Ann Transl Med.* 2014;2(12):123. doi: 10.3978/j.issn.2305-5839.2014.08.14.
 77. Witmer AN, Blaauwgeers HG, Weich HA, Alitalo K, Vrensen GFJM, Schlingemann RO. Altered expression patterns of VEGF receptors in human diabetic retina and in experimental VEGF-induced retinopathy in monkey. *Invest Ophthalmol Vis Sci.* 2002;43(3):849-57. PMID: 11867607.
 78. Markovic-Mueller S, Stutfeld E, Asthana M, Weinert T, Bliven S, Goldie KN, et al. Structure of the Full-length VEGFR-1 Extracellular Domain in Complex with VEGF-A. *Structure.* 2017;25(2):341-52. doi: 10.1016/j.str.2016.12.012.
 79. Tarallo V, De Falco S. The vascular endothelial growth factors and receptors family: Up to now the only target for anti-angiogenesis therapy. *Int J Biochem Cell Biol.* 2015;64:185-9. doi: 10.1016/j.biocel.2015.04.008.
 80. Smith GA, Fearnley GW, Harrison MA, Tomlinson DC, Wheatcroft SB, Ponnambalam S. Vascular endothelial growth factors: multitasking functionality in metabolism, health and disease. *J Inherit Metab Dis.* 2015;38(4):753-63. doi: 10.1007/s10545-015-9838-4.
 81. Caporale A, Martin AD, Capasso D, Focà G, Sandomenico A, D'Andrea LD, et al. Short PlGF-derived peptides bind VEGFR-1 and VEGFR-2 in vitro and on the surface of endothelial cells. *J Pept Sci.* 2019;25(5):e3146. doi: 10.1002/psc.3146.
 82. Carmeliet P, Moons L, Lutun A, Vincenti V, Compernelle V, De Mol M, et al. Synergism between vascular endothelial growth factor and placental growth factor contributes to angiogenesis and plasma extravasation in pathological conditions. *Nat Med.* 2001;7(5):575-83. doi: 10.1038/87904.
 83. Autiero M, Waltenberger J, Communi D, Kranz A, Moons L, Lambrechts D, et al. Role of PlGF in the intra- and intermolecular cross talk between the VEGF receptors Flt1 and Flk1. *Nat Med.* 2003;9(7):936-43. doi: 10.1038/nm884.
 84. Adams RH, Alitalo K. Molecular regulation of angiogenesis and lymphangiogenesis. *Nat Rev Mol Cell Biol.* 2007;8(6):464-78. doi: 10.1038/nrm2183.
 85. Nguyen QD, De Falco S, Behar-Cohen F, Lam WC, Li X, Reichhart N, et al. Placental growth factor and its potential role in diabetic retinopathy and other ocular neovascular diseases. *Acta Ophthalmol.* 2018; 96(1):e1-e9. doi: 10.1111/aos.13325.
 86. Tremolada G, Del Turco C, Lattanzio R, Maestroni S, Maestroni A, Bandello F, et al. The role of angiogenesis in the development of proliferative diabetic retinopathy: Impact of intravitreal anti-VEGF treatment. *Exp Diabetes Res.* 2012;2012:728325. doi: 10.1155/2012/728325.
 87. Ciulla TA, Amador AG, Zinman B. Diabetic retinopathy and diabetic macular edema: Pathophysiology, screening, and novel therapies. *Diabetes Care.* 2003;26(9):2653-64. doi: 10.2337/diacare.26.9.2653.
 88. Roldán-Pallarés M, Rollín R, Martínez-Montero JC, Fernández-Cruz A, Bravo-Llata C,

- Fernández-Durango R. Immunoreactive endothelin-1 in the vitreous humor and epiretinal membranes of patients with proliferative diabetic retinopathy. *Retina*. 2007;27(2):222-35. doi: 10.1097/01.iae.0000231376.76601.40.
89. Chang W, Lajko M, Fawzi AA. Endothelin-1 is associated with fibrosis in proliferative diabetic retinopathy membranes. *PLoS ONE*. 2018;13(1):e0191285. doi: 10.1371/journal.pone.0191285
 90. Friedlander M. Fibrosis and diseases of the eye. *J Clin Invest*. 2007;117(3):576-86. doi: 10.1172/JCI31030.
 91. Roy S, Amin S, Roy S. Retinal fibrosis in diabetic retinopathy. *Exp Eye Res*. 2016;142:71-5. doi: 10.1016/j.exer.2015.04.004.
 92. Guidry C. The role of Müller cells in fibrocontractive retinal disorders. *Prog Ret Eye Res*. 2005;24(1):75-86. doi: 10.1016/j.preteyeres.2004.07.001.
 93. Bringmann A, Reichenbach A. Role of Muller cells in retinal degenerations. *Front Biosci*. 2001;6:E72-92. doi: 10.2741/bringman.
 94. Pugh CW, Ratcliffe PJ. Regulation of angiogenesis by hypoxia: Role of the HIF system. *Nat Med*. 2003; 9(6):677-84. doi: 10.1038/nm0603-677.
 95. Smith LEH. Pathogenesis of retinopathy of prematurity. *Semin Neonatol*. 2003;8(6):469-73. doi: 10.1016/S1084-2756(03)00119-2.
 96. Wu YC, Chang CY, Kao A, Hsi B, Lee SH, Chen YH, et al. Hypoxia-induced retinal neovascularization in zebrafish embryos: A potential model of retinopathy of prematurity. *PLoS ONE*. 2015;10(5):e0126750. doi: 10.1371/journal.pone.0126750.
 97. Hinton DR, He S, Jin ML, Barron E, Ryan SJ. Novel growth factors involved in the pathogenesis of proliferative vitreoretinopathy. *Eye*. 2002;16(4):422-8. doi: 10.1038/sj.eye.6700190.
 98. Cui JZ, Chiu A, Maberley D, Ma P, Samad A, Matsubara JA. Stage specificity of novel growth factor expression during development of proliferative vitreoretinopathy. *Eye*. 2007;21(2):200-8. doi: 10.1038/sj.eye.6702169.
 99. Kuiper EJ, Van Nieuwenhoven FA, de Smet MD, van Meurs JC, Tanck MW, Oliver N, et al. The angio-fibrotic switch of VEGF and CTGF in proliferative diabetic retinopathy. *PLoS ONE*. 2008;3(7):e2675. doi: 10.1371/journal.pone.0002675.
 100. Tikellis C, Cooper ME, Twigg SM, Burns WC, Tolcos M. Connective Tissue Growth Factor Is Up-Regulated in the Diabetic Retina: Amelioration by Angiotensin-Converting Enzyme Inhibition. *Endocrinol*. 2004;145(2):860-6. doi: 10.1210/en.2003-0967.
 101. Kuiper EJ, Hughes JM, Van Geest RJ, Vogels IMC, Goldschmeding R, Van Noorden CJF, et al. Effect of VEGF-A on expression of profibrotic growth factor and extracellular matrix genes in the retina. *Invest Ophthalmol Vis Sci*. 2007;48(9):4267-76.
 102. Whitcup SM, Cidlowski JA, Csaky KG, Ambati J. Pharmacology of corticosteroids for diabetic macular edema. *Invest Ophthalmol Vis Sci*. 2018;59(1):1-12. doi: 10.1167/iovs.17-22259.
 103. Bucolo C, Gozzo L, Longo L, Mansueto S, Vitale DC, Drago F. Long-term efficacy and safety profile of multiple injections of intravitreal dexamethasone implant to manage diabetic macular edema: A systematic review of real-world studies. *J Pharmacol Sci*. 2018;138(4):219-32. doi: 10.1016/j.jphs.2018.11.001.
 104. Bucolo C, Drago F, Lin L-R, Reddy VN. Neuroactive steroids protect retinal pigment epithelium against oxidative stress. *Neuroreport*. 2005;16(11):1203-7. doi: 10.1097/00001756-200508010-00014.
 105. Bucolo C, Drago F. Neuroactive steroids protect retinal tissue through sigma1 receptors. *Basic Clin Pharmacol Toxicol*. 2007;100(3):214-6. doi: 10.1111/j.1742-7843.2007.00044.x.

106. Tabakcı BN, Ünlü N. Corticosteroid treatment in diabetic macular edema. *Turk J Oftalmol.* 2017;47(3):156-60. doi: 10.4274/tjo.56338.
107. Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, et al. Selection and analysis of an optimized Anti-VEGF antibody: Crystal structure of an affinity-matured Fab in complex with antigen. *J Mol Biol.* 1999;293(4):865-81. doi: 10.1006/jmbi.1999.3192.
108. Ferrara N, Damico L, Shams N, Lowman H, Kim R. Development of ranibizumab, an anti-vascular endothelial growth factor antigen binding fragment, as therapy for neovascular age-related macular degeneration. *Retina.* 2006; 26(8):859-70. doi: 10.1097/01.iae.0000242842.14624.e7.
109. Holash J, Davis S, Papadopoulos N, Croll SD, Ho L, Russell M, et al. VEGF-Trap: A VEGF blocker with potent anti-tumor effects. *Proc Nat Acad Sci USA.* 2002;99(17):11393-8. doi: 10.1073/pnas.172398299.
110. Rudge JS, Holash J, Hylton D, Russell M, Jiang S, Leidich R, et al. VEGF Trap complex formation measures production rates of VEGF, providing a biomarker for predicting efficacious angiogenic blockade. *Proc Nat Acad Sci USA.* 2007;104(47):18363-70. doi: 10.1073/pnas.0708865104.
111. Cai S, Yang Q, Li X, Zhang Y. The efficacy and safety of aflibercept and conbercept in diabetic macular edema. *Drug Des Devel Ther.* 2018;12:3471-83. doi: 10.2147/DDDT.S177192.
112. Papadopoulos N, Martin J, Ruan Q, Rafique A, Rosconi MP, Shi E, et al. Binding and neutralization of vascular endothelial growth factor (VEGF) and related ligands by VEGF Trap, ranibizumab and bevacizumab. *Angiogenesis.* 2012;15(2):171-85. doi: 10.1007/s10456-011-9249-6.
113. Lazzara F, Fidilio A, Platania CBM, Giurdanella G, Salomone S, Leggio GM, et al. Aflibercept regulates retinal inflammation elicited by high glucose via the PIGF/ERK pathway. *Biochem Pharmacol.* 2019;168:341-51. doi: 10.1016/j.bcp.2019.07.021.
114. Liu K, Song Y, Xu G, Ye J, Wu Z, Liu X, et al. Conbercept for Treatment of Neovascular Age-related Macular Degeneration: Results of the Randomized Phase 3 PHOENIX Study. *Am J Ophthalmol.* 2019;197:156-67. doi: 10.1016/j.ajo.2018.08.026.
115. Khan M, Aziz AA, Shafi NA, Abbas T, Khanani AM. Targeting Angiopoietin in Retinal Vascular Diseases: A Literature Review and Summary of Clinical Trials Involving Faricimab. *Cells.* 2020;9(8):1869. doi: 10.3390/cells9081869.
116. Heier JS, Singh RP, Wykoff CC, Csaky KG, Lai TYY, Loewenstein A, et al. THE Angiopoietin/Tie Pathway In Retinal Vascular Diseases: A Review. *Retina.* 2021;41(1):1-19. doi: 10.1097/IAE.0000000000003003.
117. Joly S, Dejda A, Rodriguez L, Sapieha P, Pernet V. Nogo-A inhibits vascular regeneration in ischemic retinopathy. *Glia.* 2018;66(10):2079-93. doi: 10.1002/glia.23462.
118. Mdzomba JB, Jordi N, Rodriguez L, Joly S, Bretzner F, Pernet V. Nogo-A inactivation improves visual plasticity and recovery after retinal injury. *Cell Death Dis.* 2018;9(7):727. doi: 10.1038/s41419-018-0780-x.
119. Baya Mdzomba J, Joly S, Rodriguez L, Dirani A, Lassiak P, Behar-Cohen F, et al. Nogo-A-targeting antibody promotes visual recovery and inhibits neuroinflammation after retinal injury. *Cell Death Dis.* 2020;11(2):101. doi: 10.1038/s41419-020-2302-x.
120. Falavarjani KG, Nguyen QD. Adverse events and complications associated with intravitreal injection of anti-VEGF agents: a review of literature. *Eye.* 2013;27(7):787-94. doi: 10.1038/eye.2013.107.
121. Csaky K, Do DV. Safety implications of vascular endothelial growth factor blockade

- for subjects receiving intravitreal anti-vascular endothelial growth factor therapies. *Am J Ophthalmol.* 2009;148(5):647-56. doi: 10.1016/j.ajo.2009.06.014.
122. Tolentino M. Systemic and ocular safety of intravitreal anti-VEGF therapies for ocular neovascular disease. *Survey Ophthalmol.* 2011;56(2):95-113. doi: 10.1016/j.survophthal.2010.08.006.
 123. Kurihara T, Westenskow PD, Bravo S, Aguilar E, Friedlander M. Targeted deletion of Vegfa in adult mice induces vision loss. *J Clin Invest.* 2012;122(11):4213-7. doi: 10.1172/JCI65157.
 124. De Falco S, Gigante B, Persico MG. Structure and function of placental growth factor. *Trends Cardiovasc Med.* 2002;12(6):241-6. doi: 10.1016/s1050-1738(02)00168-8.
 125. Rakic JM, Lambert V, Devy L, Lutun A, Carmeliet P, Claes C, et al. Placental growth factor, a member of the VEGF family, contributes to the development of choroidal neovascularization. *Invest Ophthalmol Vis Sci.* 2003;44(7):3186-93. doi: 10.1167/iovs.02-1092.
 126. Fischer C, Mazzone M, Jonckx B, Carmeliet P. FLT1 and its ligands VEGFB and PlGF: Drug targets for anti-angiogenic therapy? *Nat Rev Cancer.* 2008;8(12):942-56. doi: 10.1038/nrc2524.
 127. Tarallo V, Tudisco L, De Falco S. A placenta growth factor 2 variant acts as dominant negative of vascular endothelial growth factor A by heterodimerization mechanism. *Am J Cancer Res.* 2011;1(2):265-74. PMID: 21969185.
 128. De Falco S. The discovery of placenta growth factor and its biological activity. *Exp Mol Med.* 2012;44(1):1-9. doi: 10.3858/emm.2012.44.1.025.
 129. Apicella I, Cicatiello V, Acampora D, Tarallo V, De Falco S. Full Functional Knockout of Placental Growth Factor by Knockin with an Inactive Variant Able to Heterodimerize with VEGF-A. *Cell Reports.* 2018; 23(12):3635-3646. doi: 10.1016/j.celrep.2018.05.067.
 130. Cudmore MJ, Hewett PW, Ahmad S, Wang KQ, Cai M, Al-Ani B, et al. The role of heterodimerization between VEGFR-1 and VEGFR-2 in the regulation of endothelial cell homeostasis. *Nat Commun.* 2012;3:972. doi: 10.1038/ncomms1977.
 131. Tarallo V, Vesci L, Capasso O, Esposito MT, Riccioni T, Pastore L, et al. A placental growth factor variant unable to recognize Vascular Endothelial Growth Factor (VEGF) receptor-1 inhibits VEGF-dependent tumor angiogenesis via heterodimerization. *Cancer Res.* 2010;70(5):1804-13. doi: 10.1158/0008-5472.CAN-09-2609.
 132. Fischer C, Jonckx B, Mazzone M, Zachigna S, Loges S, Pattarini L, et al. Anti-PlGF Inhibits Growth of VEGF(R)-Inhibitor-Resistant Tumors without Affecting Healthy Vessels. *Cell.* 2007;131(3):463-75. doi: 10.1016/j.cell.2007.08.038.
 133. Gelfand M V, Hagan N, Tata A, Oh WJ, Lacoste B, Kang KT, et al. Neuropilin-1 functions as a VEGFR2 co-receptor to guide developmental angiogenesis independent of ligand binding. *eLife.* 2014;3:e03720. doi: 10.7554/eLife.03720.
 134. Lanahan A, Zhang X, Fantin A, Zhuang Z, Rivera-Molina F, Speichinger K, et al. The Neuropilin 1 Cytoplasmic Domain Is Required for VEGF-A-Dependent Arteriogenesis. *Dev Cell.* 2013;25(2):156-68. doi: 10.1016/j.devcel.2013.03.019.
 135. Lazzara F, Trotta MC, Platania CBM, D'Amico M, Petrillo F, Galdiero M, et al. Stabilization of HIF-1 α in Human Retinal Endothelial Cells Modulates Expression of miRNAs and Proangiogenic Growth Factors. *Front Pharmacol.* 2020;11:1063. doi: 10.3389/fphar.2020.01063.
 136. Tudisco L, Ragione F Della, Tarallo V, Apicella I, D'Esposito M, Matarazzo MR, et al. Epigenetic control of hypoxia inducible factor-1 α -dependent expression of placental growth factor in hypoxic conditions. *Epigenetics.* 2014;9:600-10. doi: 10.4161/epi.27835.

137. Yamashita T, Ohneda K, Nagano M, Miyoshi C, Kaneko N, Miwa Y, et al. Hypoxia-inducible transcription factor-2 α in endothelial cells regulates tumor neovascularization through activation of ephrin A1. *J Biol Chem*. 2008;283(27):18926-36.
138. Martinez B, Peplow P. MicroRNAs as biomarkers of diabetic retinopathy and disease progression. *Neural Regenes Res*. 2019;14(11):1858-69. doi: 10.4103/1673-5374.259602.
139. Platania CBM, Maisto R, Trotta MC, D'Amico M, Rossi S, Gesualdo C, et al. Retinal and circulating miRNA expression patterns in diabetic retinopathy: An in silico and in vivo approach. *Br J Pharmacol*. 2019;176:2179-94. doi: 10.1111/bph.14665.
140. Santovito D, Toto L, De Nardis V, Marcan-tonio P, D'Aloisio R, Mastropasqua A, et al. Plasma microRNA signature associated with retinopathy in patients with type 2 diabetes. *Sci Rep*. 2021;11(1):4136. doi: 10.1038/s41598-021-83047-w.
141. Arjamaa O, Nikinmaa M. Oxygen-dependent diseases in the retina: Role of hypoxia-inducible factors. *Exp Eye Res*. 2006;83:473-83. doi: 10.1016/j.exer.2006.01.016.
142. B. Arden G, Sivaprasad S. Hypoxia and Oxidative Stress in the Causation of Diabetic Retinopathy. *Curr Diabetes Rev*. 2012;7:291-304. doi: 10.2174/157339911797415620.
143. Kurihara T, Westenskow PD, Friedlander M. Hypoxia-inducible factor (HIF)/vascular endothelial growth factor (VEGF) signaling in the retina. *Adv Exp Med Biol*. 2014;801:275-81. doi: 10.1007/978-1-4614-3209-8_35.
144. Li T, Hu J, Gao F, Du X, Chen Y, Wu Q. Transcription factors regulate GPR91-mediated expression of VEGF in hypoxia-induced retinopathy. *Sci Rep*. 2017;7:45807. doi: 10.1038/srep45807.
145. Shao D, He S, Ye Z, Zhu X, Sun W, Fu W, et al. Identification of potential molecular targets associated with proliferative diabetic retinopathy. *BMC Ophthalmol*. 2020;20(1):143. doi: 10.1186/s12886-020-01381-5.
146. Liu X, Zhang Y, Liang H, Zhang Y, Xu Y. microRNA-499-3p inhibits proliferation and promotes apoptosis of retinal cells in diabetic retinopathy through activation of the TLR4 signaling pathway by targeting IFNA2. *Gene*. 2020;741:144539. doi: 10.1016/j.gene.2020.144539.
147. Tang X, Dai Y, Wang X, Zeng J, Li G. MicroRNA-27a protects retinal pigment epithelial cells under high glucose conditions by targeting TLR4. *Exp Ther Med*. 2018;16(1):452-8. doi: 10.3892/etm.2018.6150.
148. Ye E-A, Steinle JJ. miR-146a Attenuates Inflammatory Pathways Mediated by TLR4/NF- κ B and TNF α to Protect Primary Human Retinal Microvascular Endothelial Cells Grown in High Glucose. *Mediators Inflamm*. 2016;2016:3958453. doi: 10.1155/2016/3958453.
149. Fletcher EL, Wang AY, Jobling AI, Rutar M V, Greferath U, Gu B, et al. Targeting P2X7 receptors as a means for treating retinal disease. *Drug Discov Today*. 2019;24(8):1598-605. doi: 10.1016/j.drudis.2019.03.029.
150. Bianca Maria Platania C, Drago F, Bucolo C. The P2X7 receptor as a new pharmacological target for retinal diseases. *Biochem Pharmacol*. 2022;114942. doi: 10.1016/j.bcp.2022.114942.
151. Tassetto M, Scialdone A, Solini A, Di Virgilio F. The P2X7 Receptor: A Promising Pharmacological Target in Diabetic Retinopathy. *Int J Mol Sci*. 2021;22(13). doi: 10.3390/ijms22137110.
152. Calzaferri F, Ruiz-Ruiz C, de Diego AMG, de Pascual R, Méndez-López I, Cano-Abad MF, et al. The purinergic P2X7 receptor as a potential drug target to combat neuroinflammation in neurodegenerative diseases. *Med Res Rev*. 2020;40(6):2427-65. doi: 10.1002/med.21710.

153. Pérez de Lara MJ, Avilés-Trigueros M, Guzmán-Aránguez A, Valiente-Soriano FJ, de la Villa P, Vidal-Sanz M, et al. Potential role of P2X7 receptor in neurodegenerative processes in a murine model of glaucoma. *Brain Res Bull.* 2019;150:61-74. doi: 10.1016/j.brainresbull.2019.05.006.
154. Romano GL, Amato R, Lazzara F, Porciatti V, Chou TH, Drago F, et al. P2X7 receptor antagonism preserves retinal ganglion cells in glaucomatous mice. *Biochem Pharmacol.* 2020;180:114199. doi: 10.1016/j.bcp.2020.114199.
155. Platania CBM, Giurdanella G, Di Paola L, Leggio GM, Drago F, Salomone S, et al. P2X7 receptor antagonism: Implications in diabetic retinopathy. *Biochem Pharmacol.* 2017;138:130-9. doi: 10.1016/j.bcp.2017.05.001.
156. Platania CBM, Lazzara F, Fidilio A, Fresta CG, Conti F, Giurdanella G, et al. Blood-retinal barrier protection against high glucose damage: The role of P2X7 receptor. *Biochem Pharmacol.* 2019;168:249-58. doi: 10.1016/j.bcp.2019.07.010.
157. Fresta CG, Caruso G, Fidilio A, Platania CBM, Musso N, Caraci F, et al. Dihydro-tanshinone, a Natural Diterpenoid, Preserves Blood-Retinal Barrier Integrity via P2X7 Receptor. *Int J Mol Sci.* 2020;21(23). doi: 10.3390/ijms21239305.
158. Clapp C, Diaz-Lezama N, Adan-Castro E, Ramirez-Hernandez G, Moreno-Carranza B, Sarti AC, et al. Pharmacological blockade of the P2X7 receptor reverses retinal damage in a rat model of type 1 diabetes. *Acta Diabetol.* 2019;56(9):1031-6. doi: 10.1007/s00592-019-01343-4.